

VVR® Fast HiFi DNA Polymerase





Fast HiFi DNA Polymerase is a novel proofreading DNA polymerase. Fast HiFi DNA Polymerase is composed of a novel chimeric DNA polymerase with Archaeal ancestry, fused to a processivity-enhancing DNA binding domain. Alongside very fast and robust amplification of complex and long targets, Fast HiFi DNA Polymerase displays a high fidelity ensuring accurate amplification.

Fast HiFi DNA Polymerase is well suited for PCR experiments that require amplification with very low error rates, such as cloning/sub-cloning, NGS applications, SNP analysis and mutagenesis.

FEATURES:

- High Fidelity: > 60x Taq fidelityHigh elongation rate: 10 sec/kb
- Long range amplification: 18 kb for human gDNA
- 3'→5' proofreading exonuclease activity

HIGH FIDELITY

Fidelity values for Fast HiFi DNA Polymerase, AccuPol DNA Polymerase, two well-recognized high fidelity DNA polymerases P and Q and Taq DNA Polymerase were determined through a novel NGS-based analysis of nucleotide misincorporation during PCR. Initially, PCR amplification was performed on a ~ 200 bp synthetic DNA target, generating PCR products for each of the tested polymerases (using recommended setup conditions). Each product was purified and NGS-prepped, followed by sequencing using the MiSeq sequencing platform. In total, over 100 million reads were generated, with an average dataset size of 6 million reads. The substitution rate (error rate) was determined at each position within the DNA target (Figure 2) and subsequently summarized to determine an error rate of the entire target (Figure 1).

The error rates found for Fast HiFi polymerase and the high fidelity DNA polymerases P and Q were below the detection limit of this method, indicating that these polymerases generated very few substitution errors. The detection limit is estimated to be 8.4 x 10-6 errors per base per doubling, which corresponds to around 60x the fidelity of Taq DNA Polymerase. The error rates determined here may not be comparable with other error rates found in the literature due to technical and methodical differences.

	Error ratea	
Taq	5 x 10 ⁻⁴ (± 4,3 x 10 ⁻⁶)	
AccuPOL	1.1 x 10 ⁻⁴ (± 2.9 x 10 ⁻⁵)	
Fast HiFi	Below detection limit ^b	
Р	Below detection limit ^b	
Q	Below detection limit ^b	

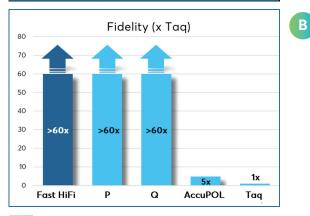
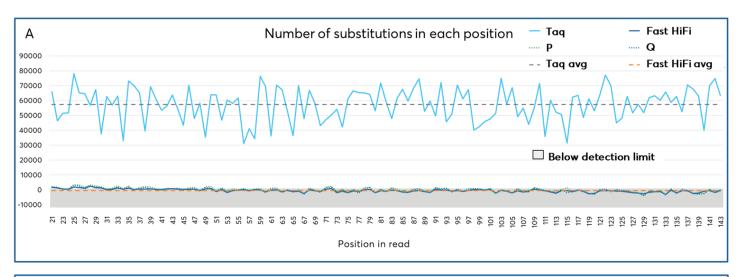


FIGURE 1: Error rates and corresponding fidelity values. **A:** Errors per base per doubling. Standard deviations are given in brackets. **B:** Fidelity values for Fast HiFi Polymerase, AccuPOL DNA Polymerase and high fidelity DNA polymerases P and Q were compared to the fidelity values of Taq DNA Polymerase (1x). * The presented error rates may not be comparable to those presented in other literature due to technical and methodical differences. * Error rates were below the detection limit for the method. This limit is estimated to be 8.4 x 10° .



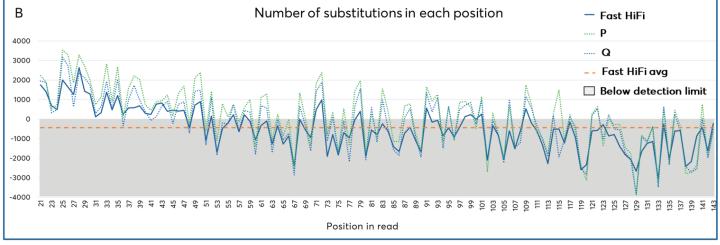


FIGURE 2: Distribution of substitutions. PCR was performed on a synthetic DNA target, using Taq DNA Polymerase, Fast HiFi polymerase and the two well-recognized high fidelity DNA polymerase P and Q. The amplified products were purified, NGS-prepped and sequenced. The number of substitutions at each target position was calculated and plotted in diagrams A and B. Diagram B magnifies the area near the detection limit. Substitutions include misincorporated nucleotides and deletions. Non-polymerase errors are subtracted from the total number of errors to reveal true polymerase errors. Non-polymerase errors include mutations caused by thermocycling-induced DNA-damage, pre-NGS sample preparation and sequencing errors. In these diagrams the average number of substitutions for Taq DNA Polymerase (Taq avg) and for Fast HiFi Polymerase (Fast HiFi avg) is also plotted.

Diagram A in Figure 2 displays the distribution profile of the substitution rate across the amplification target for Taq DNA Polymerase, Fast HiFi Polymerase and the two well-recognized high fidelity DNA polymerases Q and P. The diagram shows that the number of substitutions at each target position are much higher for Taq DNA Polymerase than for Fast HiFi Polymerase and the two high fidelity DNA polymerases P and Q. Furthermore, the number of substitutions at each target position for Fast HiFi Polymerase and the two high fidelity DNA polymerases P and Q is close to the detection limit of the

method. Diagram B magnifies the area near the detection limit, displaying more information about the number of substitutions for Fast HiFi DNA Polymerase and the high fidelity DNA polymerase P and Q.

Collectively, these diagrams show that Fast HiFi DNA Polymerase displays an extremely low numbers of substitutions. Furthermore, there is an indication that the substitution pattern of Fast HiFi Polymerase are very similar to both high fidelity DNA polymerase P and Q.

ROBUST AMPLIFICATION ON AT-RICH TO GC-RICH DNA TARGETS

Fast HiFi DNA Polymerase provides the user with robust and specific amplification of a variety of DNA targets with GC content ranging from $\sim 30-80$ % GC. The 5x Fast HiFi Buffer provided with the enzyme is recommended for highest fidelity and specificity. For DNA targets with a high GC content, more complex secondary structure or longer DNA targets, the addition of 1-2 M Betaine Enhancer Solution is recommended.

The PCR performance of Fast HiFi Polymerase was compared to that of high fidelity DNA Polymerases from three well-recognized

competitors Q, S and P (Figure 3.) PCR was performed on eight different human genomic targets, 400 – 800 bp in length and with GC content ranging from 29 – 78 % (Table 1). Robust amplification was observed for all targets using Fast HiFi Polymerase. High fidelity DNA polymerase Q and S provided results very similar to Fast HiFi DNA Polymerase, except on the last target with the highest GC content of 78 %. In contrary, high fidelity DNA polymerase P were not able to provide the same level of robust amplification on the DNA targets with higher GC content, under the conditions tested here.

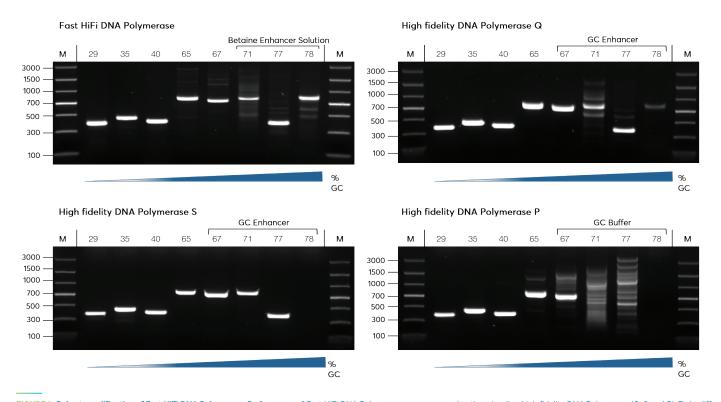


FIGURE 3: Robust amplification of Fast HiFi DNA Polymerase. Performance of Fast HiFi DNA Polymerase was compared to three leading high fidelity DNA Polymerase (Q, S and P). Eight different human genomic DNA targets, 400 – 800 bp in length and with GC content ranging from 29 – 78 %, were amplified. Amplification studies have been set up, as recommended by the manufactures. Tm Calculators of the respective competitors were used to calculate optimal annealing temperature for primers. When amplifying GC-rich targets, 2 M Betaine Enhancer Solution (Fast HiFi DNA Polymerase), GC enhancer (Competitor Q and S) or GC-rich specific PCR Buffer (competitor P) were included in the reaction mix.

% GC	Target	bp
29	CFTR-EX21	396
35	DMD19	459
40	DMD17	416
65	BAIP3	788
67	CEND	737
71	KLF14	777
77	FECH1	381
78	PO3F3	790

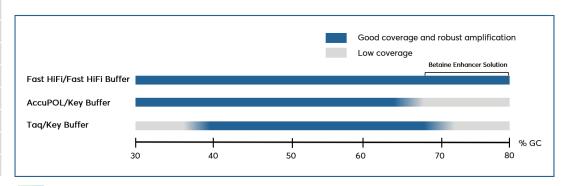


TABLE 1: DNA targets. Overview of the eight genomic DNA targets used for the amplification study despicted in figure 4. This table shows the the GC content (% GC), target names and their respective lenghts (bp).

FIGURE 4: Illustration of the coverage of Fast HiFi DNA Polymerase. 5x Fast HiFi Buffer supports robust amplification of DNA targets with a GC content ranging from ~ 30 ~ 80 %. The addition of 2M Betaine Enhancer solution supports amplification of DNA targets with high GC content. The coverage of Fast HiFi DNA Polymerase is illustrated against the coverage of AccuPol DNA polymerase and Taq DNA Polymerase when using the 10x Key Buffer.

LONG RANGE AMPLIFICATION

Fast HiFi DNA Polymerase provides the user with the ability to amplify a broad range of DNA targets from short and up to 18 kb for human genomic DNA (Figure 5).

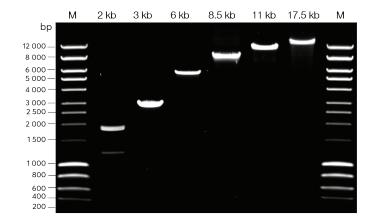


FIGURE 5: Fast HiFi DNA Polymerase enables long range amplification. Six different targets of human genomic DNA ranging from 2 kb and up to 17.5 kb was used in this study. Amplicon sizes are indicated at the top of the gel. DNA marker (M).

APPLICATIONS:

- Cloning/sub-cloning
- Long range amplification
- NGS applications
- Mutagenesis
- Gene expression
- Construction of libraries
- SNP analysis

Ordering information:

Product	Size	Cat. No.
Fast HiFi DNA Polymerase 2 U/µl	100 units	733-2880
Incl. 5x Fast HiFi Buffer and 25 mM MgCl2	500 units	733-2881
	1000 units	733-2882
	2500 units	733-2883
Fast HiFi DNA Polymerase 2X Master Mix	100 reactions	733-2884
1.5 mM MgCl2 final conc.	500 reactions	733-2885
	2500 reactions	733-2886
	5000 reactions	733-2887

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AUSTRIA

VWR International GmbH Graumanngasse 7 1150 Wien Tel.: +43 1 97 002 0 info.at@vwr.com

BELGIUM

VWR International bv Researchpark Haasrode 2020 Geldenaaksebaan 464 3001 Leuven Tel.: +32 (0) 16 385 011 vwr.be@vwr.com

CZECH REPUBLIC

VWR International s. r. o. Veetee Business Park Pražská 442 CZ - 281 67 Stříbrná Skalice Tel.: +420 321 570 321 info.cz@vwr.com

DENMARK

VWR International A/S Tobaksvejen 21 2860 Søborg Tel.: +45 43 86 87 88 info.dk@vwr.com

FINLAND

VWR International Oy Valimotie 9 00380 Helsinki Tel.: +358 (0) 9 80 45 51 info.fi@vwr.com

FRANCE

VWR International S.A.S. Le Périgares – Bâtiment B 201, rue Carnot 94126 Fontenay-sous-Bois cedex Tel.: 0 825 02 30 30* (national) Tel.: +33 (0) 1 45 14 85 00 (international) info.fr@vwr.com

GERMANY

VWR International GmbH Hilpertstraße 20a D - 64295 Darmstadt Tel.: 0800 702 00 07* (national) Tel.: +49 (0) 6151 3972 0 (international) info.de@vwr.com

HUNGARY

VWR International Kft. Simon László u. 4. 4034 Debrecen Tel.: +36 52 521130 info.hu@vwr.com

IRELAND

VWR International Ltd Orion Business Campus Northwest Business Park Ballycoolin Dublin 15 Tel.: +353 (0) 1 88 22 222 sales.ie@vwr.com

ITALY

VWR International S.r.l. Via San Giusto 85 20153 Milano (MI) Tel.: +39 02 3320311 info it@ywr.com

THE NETHERLANDS

VWR International B.V. Postbus 8198 1005 AD Amsterdam Tel.: +31 (0) 20 4808 400 info.nl@vwr.com

NORWAY

VWR International AS Brynsalleen 4, 0667 Oslo Tel.: +47 22 90 00 00 info.no@vwr.com

POLAND

VWR International Sp. z o.o. Limbowa 5 80-175 Gdansk Tel.: +48 58 32 38 200 info.pl@vwr.com

PORTUGAL

VWR International - Material de Laboratório, Lda Centro Empresarial de Alfragide Rua da Indústria, nº 6 2610-088 Amadora Tel.: +351 21 3600 770 info.pt@vwr.com

SPAIN

VWR International Eurolab S.L.U. C/Tecnología 5-17 A-7 Llinars Park 08450 - Llinars del Vallès Barcelona Tel.: +34 902 222 897 info.es@vwr.com

SWEDEN

VWR International AB Fagerstagatan 18a 163 94 Stockholm Tel.: +46 (0) 8 621 34 00 kundservice.se@vwr.com

SWITZERLAND

VWR International GmbH Lerzenstrasse 16/18 8953 Dietikon Tel.: +41 (0) 44 745 13 13 info.ch@vwr.com

UK

VWR International Ltd Customer Service Centre Hunter Boulevard - Magna Park Lutterworth Leicestershire LE17 4XN Tel.: +44 (0) 800 22 33 44 uksales@vwr.com

CHINA

VWR (Shanghai) Co., Ltd Bld.No.1, No.3728 Jinke Rd, Pudong New District Shanghai, 201203- China Tel.: 400 821 8006 info_china@vwr.com

ÍNDIA

Avantor Performance Materials India Limited 17th Floor, Building No. 5, Tower C DLF Cyber City Phase – III Gurgaon - 122002, Haryana Tel.: +91-1244-65-6700 help@avantorinc.com

KOREA

VWR International ~ 17, Daehak 4-ro, Yeongtong-gu, Suwon-si, Gyeonggi-do Tel.: +82 31 645 7256 saleskorea@avantorsciences.com

MIDDLE EAST & AFRICA

VWR International FZ-LLC Office 203, DSP Lab Complex, Dubai Science Park, Dubai, United Arab Emirates Tel: +971 4 5573271 Info.mea@vwr.com

SINGAPORE

VWR Singapore Pte Ltd 18 Gul Drive Singapore 629468 Tel: +65 6505 0760 sales.sa@vwr.com

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